REVIEW

Aluminum Toxicity and Tolerance Mechanism in Cereals and Legumes - A Review

Kodithuwakku Kankanange Indika Upali Arunakumara · Buddhi Charana Walpola · Min-Ho Yoon

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Abstract Aluminum (Al), the third most abundant element overall, after silicon and oxygen, is found virtually in all food, air, soil and water. Under acidic conditions, Al is solubilized into $[Al(H_2O)_6]^{3+}$, which is highly toxic to many plant species. Present paper reviewed Al toxicity and tolerance mechanism in agricultural plants with special reference to cereals and legumes. Even at micromolar concentrations, cell division in the root tip meristem in sensitive plants is quickly inhibited by Al³⁺. Expressing the species-dependent manner of response, growth enhancement at low concentrations of Al³⁺ is also reported from some plants such as soybean. Plasma membrane can be identified as the primary target of Al toxicity, where production of higher reactive oxygen species and higher fatty acid peroxidation has been observed due to alteration of plasma membrane integrity. Though, toxicity and the mechanisms attributed to Al-resistance are extremely complex phenomena, exclusion is widely accepted as the key mechanism involved in detoxifying Al3+. Exudation of chelating ligands, formation of pH barrier at the rhizosphere or at root apoplasm, cell wall immobilization, selective permeability of the plasma membrane, and Al efflux have been proposed as the possible mechanisms for Al exclusion. Al-induced exudation in cereals and legumes is dominated by citrate, malate, and oxalate in varying degrees depending on the species and/or cultivar. Apart from sensitive cultivars, moderately tolerant or tolerant cultivars can also be distinguished from various kinds of cereals and legumes. However, reliable techniques for screening such resistant genotypes have not been developed for any economically important crops.

Keywords exudation · ligands · metabolism · tolerance · toxicity

Aluminum toxicity

In terms of relative abundance, aluminum (Al) is the third element after oxygen and silicon and the most abundant metal present in the earth's crust (Ma and Furukawa, 2003; Matsumoto and Motoda, 2012). It belongs to the non-essential category of metals, thus does not exert any known function in plant metabolism (Wang and Kao, 2004). However, the metal is considered to be a major growth-limiting factor particularly in acid soils (pH <5.0), which are estimated to be approximately 30-40% of arable lands in the world (Lilienfein et al., 2003). Depending on pH, Al exists in a number of different forms in the soil (Wang et al., 2006). Under acidic conditions, Al is solubilized into $[Al(H_2O)_6]^{3+}$, generally referred to as Al³⁺, which is highly toxic to many plant species (Éva Darkó et al., 2004). Despite the fact that many plants find it difficult to withstand against the Al toxicity, its most common forms i.e. oxides and aluminosilicates are harmless to plants (Ma and Furukawa, 2003; Wang and Kao, 2004).

In sensitive plants, cell division in the root tip meristem is quickly inhibited by Al^{3+} , thus root elongation is immediately affected (Gunsé et al., 2003; Doncheva et al., 2005; Meda and Furlani, 2005). This occurs because the apex of the root (root cap, meristem, and elongation zone) could accumulate comparatively higher amount of Al^{3+} than the other parts of the root (Zheng and Yang, 2005). Increased rigidity of the double helix of DNA caused by accumulated Al^{3+} results in poor cell division in root tip meristem (Meriga et al., 2010). In many plants, inhibition of root elongation has been reported within a few minutes even at micromolar concentrations of Al^{3+} (Matsumoto and Motoda, 2012), though enhanced root activity at low concentrations of Al^{3+} has also been reported in soybean (Yu et al., 2011). This might be due to Al-induced increasing activity of the apical meristem. In

K. K. I. U. Arunakumara

Department of Crop Science, Faculty of Agriculture, University of Ruhuna, Mapalana, Kamburupitiya, Sri Lanka

B. C. Walpola · M.-H. Yoon (🖂)

Department of Bio-Environmental Chemistry, College of Agriculture and Life Sciences, Chungnam National University, Daejeon, 305-764, Republic of Korea

E-mail: mhyoon@cnu.ac.kr

some cases, increased susceptibility to drought stress, lodging, and nutrient deficiencies are also reported from affected plants (Sun et al., 2010). The early signs of Al toxicity could be observed on the cells of the epidermis or outer cortex, which may be due to direct interaction of Al3+ with the plasma membrane (Barceló and Poschenrieder, 2002). Based on the results of several investigations, plasma membrane can be identified as the primary target of Al toxicity (Gunsé et al., 2003; Ahn and Matsumoto, 2006; Silva, 2012). According to Peixoto et al. (2001), production of higher reactive oxygen species and higher fatty acid peroxidation of the plasma membrane in Al-sensitive cultivars is due to deteriorated performance of affected plasma membrane. Therefore, growth reduction and alteration of plasma membrane integrity are the immediate consequences of binding of Al3+ to cell membrane components. Discovering another feature of Al toxicity, Sun et al. (2007; 2010) have reported that ethylene and auxin are also involved in inhibition of root elongation under Al stress. In summarizing, Tables 1 and 2 can be considered as the possible consequences of Al toxicity, which may vary depending on the species and the cultivar.

- Inhibition of cell division, root elongation and plant growth (Doncheva et al., 2005)
- Alteration of nutrient availability in the rhizosphere (Poschenrieder et al., 2008)
- Accumulation of certain metabolites (Peixoto et al., 2001; Amenós et al., 2009)
- Behavioral changes in many key enzymes (Zhou et al., 2009)
- Low P availability which can result in decreased photosynthetic efficiency and low dry matter production (Chaudhary et al., 2008)
- Increased susceptibility to drought stress, lodging, and nutrient deficiencies (Sun et al., 2010)
- High affinity of cationic Al for plant cell walls, membranes and metabolites is known to be the plants' quick response to Al toxicity (Kochian et al., 2005; Hiradate et al., 2007), which is often assessed with the use of different plant indicators such as inhibition of cell division and cell expansion (Ma et al., 2004; Horst et al., 2010), fine root development (Zobel and Kinraide, 2007), root elongation (Rangel et al., 2009), alteration of nutrient acquisition processes, and cell homeostasis (Purcino et al., 2003; Garzon et al., 2011), as well as changes in cytosolic Ca²⁺, pH and K⁺ levels, and plasma membrane polarization (Vanguelova et al., 2007; Bose et al., 2010), accumulation of callose (Hirano and Brunner, 2006), Al specific tissue staining (Tolrà et al., 2007).

Sensitivity and Tolerance

Al-sensitive and Al-resistant species/cultivars have been widely investigated and several studies revealed that Al phytotoxicity and the mechanisms attributed to Al-resistance are extremely complex phenomena (Matsumoto, 2000; Ciamporová, 2002; Éva Darkó et al., 2004). In mitigating the adverse impacts of Al toxicity, plants use different strategies, of which exclusion is the widely accepted mechanism (Delhaize et al., 2007; Garzon et al., 2011). Exclusion is often proceeded with one or combination of following processes; exudation of chelating ligands, formation of pH barrier at the rhizosphere or at root apoplasm, cell wall immobilization, selective permeability of the plasma membrane, and Al efflux (Éva Darkó et al., 2004; Wang et al., 2006; Delhaize et al., 2007; Tolra et al., 2009).

Among the above, exclusion of toxic Al³⁺ through chelating ligands is, however, the dominant detoxification mechanism for many plant species (Poschenrieder et al., 2005; Maron et al., 2010). In fulfilling this, organic acids such as citrate, malate, and oxalate are reported to be exudated at the rhizosphere and the apoplast (Ryan et al., 2001; Poschenrieder et al., 2005). In fact, exudation of malate, citrate and/or oxalate by root tips is recognized to be the key factor in Al-tolerance of many species due to their high affinity for binding of Al^{3+} (Kochian et al., 2005; Ryan et al., 2011). Elevation of pH at the surrounding environment is considered to be one of the other tolerance mechanisms, because high pH reduces the Al solubility, which leads the formation of less-toxic Al species, such as Al-hydroxides and Al-phosphates (Wang et al., 2006). Exudation of organic acids from roots is also encouraged by elevation of pH at rhizosphere. However, Garzon et al. (2011) elaborating their findings of the experiment carried out with two maize varieties reported that neither rhizosphere pH changes nor Al-tolerance were clearly related to Al-induced changes in exudation of organic acids into the rhizosphere. Formation of stable complex with ionic Al is known for some of these organic acids, thus, can prevent binding of Al3+ with extra- and intracellular substances of the root (Li et al., 2000). Chelation of Al^{3+} by other forms of ligands in the cytosol (Ma et al., 2001), Al compartmentation in the vacuole, and several other possible Al³⁺ protective mechanisms are also found in the literature (Éva Darkó et al., 2004; Delhaize1 et al., 2012).

The responses of plants to Al toxicity are highly variable within the species; more importantly, responses from different genotypes of a same species can also vary (Smith et al., 2011). Investigations on the toxicity and resistance mechanisms have often been performed taking physiological and genetic basis of resistance into consideration (Kochian et al., 2005; Godbold and Brunner, 2007). However, quantitative data needed to assess the magnitude of the physiological damage are still found to be scarce (Ruf and Brunner, 2003; Nagy et al., 2004; Hirano and Brunner, 2006; Vanguelova et al., 2007). Despite the fact that substantial attention is needed to be paid on the recovery mechanism from Al-induced injuries, information on the rehabilitation process is also lacking (Kikui et al., 2007; Motoda et al., 2010; 2011). In this context, Matsumoto and Motoda (2012), in reviewing Al toxicity recovery processes in root apices, have reported that at elevated concentrations of Al³⁺, recovery is hardly noticeable, because root elongation stops completely, causing plants to begin dying. According to

Crop	Var./Culti.	Symptom/s	References
	Sikuani (tolerant)	No inhibition of root elongation No effects of Al on cytoplasmic Ca ²⁺ Increased vacuolar nitrate concentration Decreased vacuvacuolar malate concentration	Garzon et al. (2011)
	Bakero (sensitive)	Significant inhibition of root elongation Decreased cytoplasmic Ca ²⁺ in root tips Increased vacuvacuolar nitrate concentration Increased vacuvacuolar malate concentration	
Maize (Zea mays L.)	ATP SR Yellow (tolerant)	Higher root elongation Less callose production	Gunsé et al. (2000)
	HS 701 B (sensitive) Cateto (tolerant)	Poor root elongation More callose production No inhibition of root elongation Roots accumulations of caffeic acid, catechol and catechin are high	Tolra et al. (2009)
	HS16 x 36 (sensitive)	Significant inhibition of root elongation Roots accumulations of caffeic acid, catechol and catechin are poor	
	Atlas 66 (tolerant)	Enhanced exudation of malic and citric acids High rhizosphere pH	Wang et al. (2006)
Wheat (Triticum aestivum L.)	Brevor (sensitive)	Accumulation of more Al in roots No exudation of organic acids Marked inhibition of root elongation	
	Scout 66 (sensitive)	Severe reduction in cellulose synthesis Marked inhibition of root elongation	Teraoka et al. (2002)
	Xiushui 132 (tolerant)	Slight growth inhibition Reduction in chlorophyll content Increased proline and malonaldehyde contents Increased peroxidase and superoxide dismutase activities	Guo et al. (2012)
Rice (<i>Oryza sativa</i> L.)	Yongyou 8 (sensitive)	Severe growth inhibition Severe reduction in chlorophyll content Significant increased in proline and malonaldehyde contents Significant increased in peroxidase and superoxide dismutase activities	
	Suraksha (tolerant) Vikas (sensitive)	Lesser reduction in root and shoot length Higher reduction in root and shoot length Increased lipid peroxidation and higher activity of SOD and peroxidase	Meriga et al. (2010)

Matsumoto and Motoda (2012), recovery and exudation of organic acids are of steps connected to each other. Exudation of organic acids begins with Al-induced stress. Recovery of injured roots begins to appear as a consequence of toxicity reduction by means of chelation. Once recovered, re-activated roots encounter toxic A^{3+} , which then forces the re-commencement of organic acids exudation. Under certain environmental conditions, inhibition of root elongation and the recovery process come into act simultaneously or may be as repeated steps, one following the other.

Cereals and Legumes

Cereals, belong to family Poaceae, are economically the most important crops in the world, because they are the staple food sources in many nations (León-Ruiz et al., 2011). Due to widespread distribution and the diversified genetic base, cereals can now be found in all continents, regardless of latitude, soil, and climate. Cultivation of cereals in different scales under different cropping systems is found almost everywhere, in particular, with the introduction of genetically modified and enhanced varieties of

Crop	Var./Culti.	Symptom/s	References
Soybean (<i>G. ma</i> x L.)	Zhechun 2 (tolerant)	Severe reduction in root length and volume Decreased root activity Increased plasma membrane permeability Disorganized cytoplasm of root tip cells	Yu et al. (2011)
	Zhechun 3 (sensitive)	Severe reduction in root length and volume Marked decreased in root activity Marked increased in plasma membrane permeability Disorganized cytoplasm of root tip cells	
	Zhechun 2 (tolerant)	Increased inhibition of root elongation Dose-dependent pectin methylesterase activity in the root tip	Cai et al. (2011)
	Huachun 18 (sensitive)	Significantly increased inhibition of root elongation Dose-independent pectin methylesterase activity in the root tip	
Pea (P. sativam L.)	Rachana (tolerant)	Significantly reduced seed germination and seedling growth Increased lipid peroxidation High content of superoxide dismutase, catalase, ascorbate peroxidase, guaicol peroxidase, ascorbic acid and non protein thiol in root tips	Singh et al. (2011)
	Arkil (sensitive)	Significantly reduced seed germination and seedling growth Significantly increased lipid peroxidation Low content of superoxide dismutase, catalase, ascorbate peroxidase, guaicol peroxidase, ascorbic acid and non protein thiol in root tips	
Common bean (<i>P. vulgaris</i> L.)	Preto and Carioca (tolerant)	Slight inhibition of root elongation	Gunsé et al. (2003)
	Contender and Superba (sensitive)	Severe inhibition of root elongation	
	Andean (tolerant)	Slight reductions in the elongation rate of the primary roots, total root length and specific root length	Blair et al. (2009)
	Mesoamerican (sensitive)	Higher reductions in the elongation rate of the primary roots, total root length and specific root length	
	Quimbaya (tolerant)	Severe inhibition of root elongation with substantial recovery High content of cell wall pectin	Rangel et al. (2009)
	VAX-1 (sensitive)	Severe inhibition of root elongation with no recovery Low content of cell wall pectin	

Table 2 Varietal differences in three legume crops in response to Al toxicity

specific species and better-adapted hybrids (Damialis and Konstantinou, 2011). Recent study showed that over the first decade of the 21st century, leading cereal crops (maize, wheat, and rice) have contributed 80–90% of world grain production, and almost 40% of calorie intake of the world population (Van Wyk, 2005). Cereals are the only group of crops with cultivation that exceeds 20% of global land area or 61% of the total cultivated land. Wheat (*Triticum aestivum* L.), maize (*Zea mays* L.), rice (*Oryza sativa* L.), barley (*Hordeum* spp.), millet (Small-grained cereals that include a large number of different botanical species), and rye (*Secale cereal* L.) are considered to be dominant cereal crops.

Legumes (family-Fabaceae) are the third largest family of angiosperms after Orchidaceae and Asteraceae. There are about

20,000 species belong to family Fabaceae (Doyle and Luckow, 2003), and the family has a significant agricultural importance, because some key legumes such as soybean (*Glycine max* L.), pea (*Pisum sativam* L.), common bean (*Phaseolus vulgaris* L.), mungbean (*Vigna radiate* L.), peanut (*Arachis hypogeae* L.), pigeon pea (*Cajanus cajan* L.), and cowpea (*Vigna unguiculata* L.) are cultivated on 12–15% of the arable land throughout the world (Graham and Vance, 2003). Legumes also bring remarkable additional benefits, in particular their capacity to fix atmospheric nitrogen through mutualistic interactions with rhizobial soil bacteria. As to how cereals and legumes response to Al toxicity is discussed below with major emphasis given to the key economically important species.

Maize (Z. mays L.)

Maize, a widely known food crop, is gaining worldwide attention as a source of biofuel as well. When expose to Al, responses differ among different maize varieties, thus sensitive and moderately tolerant varieties can be distinguished (Meda and Furlani, 2005). Exclusion as the dominant defense mechanism against the Al toxicity in maize is often discussed. Kidd et al. (2001) reported that as in many other crops, Al-resistance in maize (Var. Sikuani) is attributed to exudation of strong Al chelating agents such as malate and citrate. However, Garzon et al. (2011) reported that Alinduced enhancement of root exudation of citrate was hardly detected in maize (Var. Sikuani and Bakero). Based on their findings, where a weak correlation between organic acid exudation and Altolerance has been observed, other contributors may also come into play a substantial role in Al-tolerance of maize as reported previously by Piñeros et al. (2005) and Tolra et al. (2009). Tolra et al. (2009) investigated Al-induced pattern of phenolic compounds of two maize varieties and concluded that, in addition to Al exclusion from the roots, higher contents of phenolic substances with a high antioxidant and antiradical activities also contribute to Al-tolerance in maize. Liu et al. (2009) also confirmed the Alinduced root tip exudation of flavonoid type phenolics in maize. However, due to high affinity for H⁺, phenolics gain lower recognition compared to organic acids in acidic soils. Kidd et al. (2001) reported that Al-induced exudation of flavonoid-type phenolics could also be contributed to silicon-mediated amelioration of Al toxicity in maize. However, even in highly tolerant varieties, Al manages to gain access into the cells as observed by Gaume et al. (2001). Therefore, once Al³⁺ has managed to enter the symplasm, internal detoxification by means of chelation and/or sequestration to less sensitive compartments is equally important (Tolra et al., 2005; Klug and Horst, 2010). As reported by Tolra et al. (2009), compounds bearing catecholate moieties are also strong ligands for Al³⁺. Detoxification of Al³⁺, which has already surpassed the exclusion barrier, is done by these catecholate moieties. Poschenrieder et al. (2005) used callose and haematoxylin staining as indicators for Al toxicity and reported that 2,4-dihydroxy-7-methoxy-1,4benzoxazin-3-one (DIMBOA) by binding Al3+ can protect the root tips of sensitive maize plants against Al toxicity in vitro. Root growth inhibition similar to many other crop species has also been reported in maize (Kochian, 2004; Li et al., 2004). As recorded in several studies, Al³⁺ possess the ability to bind to carboxyl and phosphoryl groups of the cell wall and the plasma membrane, thus specific changes in the lipid composition of the plasma membrane could also contribute in maintaining root growth under Al stress (da Silva et al., 2006). In addition to root exudation, contribution from phenolic compounds to the defense mechanism in maize is considered to be encouraging findings. Though reliable screening techniques are yet to be developed, reports available on tolerant cultivars are also important.

Wheat (T. aestivum L.)

In the case of Wheat, a leading staple food crop in the world, Alinduced inhibition of root elongation is widely known (Teraoka et al., 2002; Zakir et al., 2003). The stress can alter the ultrastructure of root cells (Xiao et al., 2003; Li et al., 2006). Alteration of the physical dimensions of the root apex in wheat has come to appear due to shortening of the root elongation zone while increasing in the diameter of the cells on the outer cortex, which ultimately results in ruptures of the epidermis (Motoda et al., 2010). Structural alterations in similar nature have also been reported in maize. As reported by Matsumoto and Motoda (2012), the rigidity of the cell wall increases with the inhibition of root elongation under Al stress. Among the defense mechanisms, concentration-dependent efflux of malate from root tips is reported in wheat also (Zhang et al., 2001), though capacity to release malate varies with the cultivar. Osawa and Matsumoto (2001) have suggested that protein phosphorylation is involved in the Al-responsive malate efflux in the wheat root apex. Darkó et al. (2004) investigated the formation of reactive oxygen species induced by Al stress in sensitive genotype and in tolerant lines developed by in vitro microspore selection. The roots of tolerant plants were found to accumulate less Al³⁺ and reactive oxygen species than sensitive plants. They concluded that among the superoxide dismutase, ascorbate peroxidase (APX), catalase (Cat), and glutathione-S-transferase (GST) enzymes induced by Al stress, catalase and GST may play an important role in the detoxification of reactive oxygen species in Al-tolerant wheat plants. Two wheat cultivars, Darab (Al-sensitive) and Maroon (Al-tolerant) grown on hydroponic solution were used in assessing growth of wheat seedlings under Al stress (Nasr et al., 2011). Results revealed that, for fresh weight of root, fresh weight of shoot, dry weight of shoot and length of root, Al concentrations and their interaction with cultivar were highly significant. They have further reported that application of malate or citrate is effective in diluting the Al toxicity in wheat seedlings. Among cereals, wheat has been employed extensively in exploring many aspects related to Al toxicity, enabling genetic engineering to engage in producing wheat cultivars with greater resistance. Therefore, continuous production of wheat even in acidic soils would be ensured in due course.

Rice (O. sativa L.)

Rice is basically considered to be an Al-tolerant crop, although the response is largely dependent upon the cultivar of concern (Guo et al., 2012). Meriga et al. (2010) investigated the effect of Al on seedlings of two rice cultivars (Vikas and Suraksha) under different pH regimes and reported that as pH reduces, reduction in plant growth, accumulation of Al^{3+} in roots, lipid peroxidation, and activity of superoxide dismutase (SOD) and peroxidase were

more pronounced in Vikas, the sensitive cultivar. Furthermore, they have observed an increasing resistant with the age of the seedlings even in sensitive cultivar. Increased activity of SOD, one of the most effective anti-oxidative enzymes, which prevents cellular damage, has been reported previously (Sharma and Dubey, 2007) also in rice seedlings in response to Al toxicity. Hydroponic experiments conducted by Guo et al. (2012) to assess the Al toxicity and P deficiency in two rice genotypes (Yongyou 8 and Xiushui 132) also revealed growth inhibition along with reduced chlorophyll content, increased proline and malonaldehyde contents, and increased peroxidase and superoxide dismutase activities in roots. Their findings can of course be considered as re-confirmation of previous findings (Ismail, 2005; Sarker and Karmoker, 2011) with some other crops such as mungbean, carrot, radish, and lentil seedlings. Based on the results of higher proline content in Al-tolerant genotype over the sensitive cultivar, Guo et al. (2012) suggested that the difference between the cultivars in response to Al toxicity could be assessed based on proline content. Well documented Al-induced inhibition of root elongation is also reported in rice (Kikui et al., 2005). Kuo and Kao (2003) and Tewari et al. (2004) reported an increased malonaldehyde (MDA) content in rice seedlings under Al and P starvation stress. Confirming these previous reports, Guo et al. (2012) observed similar response pattern indicating that lipid peroxidation occurred as a result of reactive oxygen species (ROS) formation induced by high level Al toxicity. Reports with tolerant cultivars draw key attention in the field of Al toxicity to rice. Distinguishing such cultivars based on higher proline content is further encouraging.

Soybean (G max L.)

Soybean, an important source of high quality but inexpensive protein (40%) and oil (20%) is grown in over 90 million ha throughout the world. Yu et al. (2011) studied the effect of Al on the root growth and cell ultrastructure of two soybean genotypes (Zhechun 2 and 3 respectively Al-resistant and -sensitive) and observed significant reduction in root biomass at higher concentration (90 mg/L) of Al^{3+} with a slight difference between two genotypes. However, root activity of sensitive variety was significantly lower than that of resistant. Furthermore, they have observed alterations in the cellular organization, mitochondria, vacuoles, and nuclei physiology with marked varietal differences as well as reduced root elongation and development when Al concentration in the solution was increased. Confirming the earlier findings (Kollmeier et al., 2000), root tip and prolongation zone have been identified by Yu et al. (2011) also as the locations, which underwent sever difficulties resulting immediate toxicity symptoms. Most importantly, they have stressed that under low concentrations (depending on the variety of soybean, 10-30 mg/L), Al could enhance root activity, maintain high respiration metabolism, and strengthen the absorption abilities of water and minerals, which is in agreement with Li et al. (2004). Growth enhancement in legumes under low

pH is of particular important, because the growth of rhizobia with which soybean is known to have deep symbiotic relationship could be badly affected by the acidic environment. Rhizobial growth is considered to be optimum at pH between 6.0 and 7.0 (Hungria and Vargas, 2000) and as of previous reports (Ying et al., 2005), number of nitrogen-fixing bacteria also decreases with the increase of Al concentration. In addition to the well known toxicity response, growth enhancement at low concentrations of Al^{3+} , alterations in the cellular organization and potential threat to the associated bacteria etc., could be considered as important findings of the researches with Al toxicity to soybean.

Pea (P. sativam L.)

Pea, a source of high quality protein in human diet and animal feed, is grown for its seeds. Matsumoto and Motoda (2012) elaborating the process of recovery form of pea roots exposed to Al and reported that injuries come into appear more intensively in closely located cells in the region of Al uptake, where unique structural alterations begin to appear quickly. According to them, it takes only a few seconds to reach the metal to the cell wall of epidermal cells, whereas time taken for reaching other root tissues was about one hour. Therefore, starting from epidermal cells, Alinduced injuries could be observed in outer cortex, middle cortex, inner cortex and style as reported previously by Ciamporova (2002). Singh et al. (2011) studied the morphological and biochemical characteristics of two pea varieties (Arkil and Rachana) under Al stress in soil and observed remarkably reduced seed germination and seedling growth in both the varieties. Furthermore, Rachana, the tolerant variety increased the activities of superoxide dismutase (SOD), catalase (CAT), ascorbate peroxidase (APX), guaicol peroxidase (GPX), ascorbic acid (AsA) and non protein thiol (NPSH) markedly compared to those of Arkil, the sensitive variety. Enhanced production of reactive oxygen species, a common feature of Al toxicity has been reported in Pea also. As of the reports so far published, moderately tolerant cultivars such as Rachana have been identified and available for further studies.

Common bean (P. vulgaris L.)

Common bean is considered to be the second important food legume, in particular, in developing countries. Investigations conducted with soybean, pea and common bean have been used, in part, to answer the much debated question; whether the initial toxicity display of Al is on inhibition of cell elongation or cell division. It is the present understanding that in legumes, cell elongation and cell division are both inhibited; however, responses due to inhibition of cell division are largely irreversible (Barcelo and Pochenrieder, 2002). Rangel et al. (2005) observed a positive correlation between Al-induced callose formation and inhibition of root elongation and Al contents in the root apices of common bean. Though some genotypes such as Andean and Mesoamerican have been found to display greater performance under acidic soil conditions, Rangel et al. (2005) suggested having more investigations in order to understand genotypic differences in response to Al toxicity and to develop quick screening techniques for Al resistance. Not like for other key legume species, literature on Al toxicity in common bean is still lacking in many aspects. Therefore, further investigations are needed, in particular, to distinguish varietal differences in response to Al toxicity.

Conclusion

Though the focused areas of this review have been investigated over the years, contradictory reports such as inhibition, as well as growth enhancement at low concentration of Al³⁺ are often found. This might be due to species-dependent manner of response and/ or differences in experimental material and methodology used. In addition, the vast majority of the investigations have been aimed at elucidating the effect of Al on a particular parameter of the plant; thus comprehensive understanding on as to how Al affects overall performance of plants has yet to be elucidated. It is necessary to conduct, more studies under field conditions to understand the process of recovery from Al toxicity, because findings of such investigations would have undeniable practical significance. Due to little attention so far paid on the recovery process, development of appropriate agronomic practices to ensure speedy recovery from Al toxicity is also found to be lacking. Development of reliable screening techniques for resistant genotypes would be another area to be worked on. However, taking the species-dependent nature of response into consideration, a series of researches are needed to identify dependable parameters for use in such screening methods.

References

- Ahn SJ and Matsumoto H (2006) The role of the plasma membrane in the response of plant roots to aluminum toxicity. *Plant Signal Behav* 1, 37– 45.
- Amenós M, Corrales I, Poschenrieder C, Illéš P, and Baluška F (2009) Different effects of aluminum on the actin cytoskeleton and brefeldin asensitive vesicle recycling in root apex cells of two maize varieties differing in root elongation rate and aluminum tolerance. *Plant Cell Physiol* **50**, 528–40.
- Barcelo J and Pochenrieder C (2002) Fast root growth response, root exudates, and internal detoxification as clues to the mechanisms of aluminum toxicity and resistance, a review. *Environ Exp Bot* 48, 75–92.
- Blair MW, López-Marín HD, and Rao MI (2009) Identification of aluminum resistant Andean common bean (*Phaseolus vulgaris* L.) genotypes. *Braz J Plant Physiol* 21, 291–300.
- Bose J, Babourina O, Shabala S, and Rengel Z (2010) Aluminum-induced ion transport in Arabidopsis: the relationship between Al tolerance and root ion flux. J Exp Bot 61, 3163–75.
- Cai MZ, Wang FM, Li RF, Zhang SN, Wang N, and Xu GD (2011) Response and tolerance of root border cells to aluminum toxicity in soybean

seedlings. J Inorg Biochem 105, 966-71.

- Chaudhary ML, Adu-Gyamfi JJ, Saneoka H, Nguyen NT, Suwa R, Kanai S et al. (2008) The effect of phosphorus deficiency on nutrient uptake, nitrogen fixation and photosynthetic rate in mashbean, mungbean and soybean. *Acta Physiol Plant* **30**, 537–44.
- Ciamporova M (2002) Morphological and structural response of plant roots to aluminum at organ, tissue and cellular levels. *Biol Plant* 45, 161–71.
- Damialis A and Konstantinou GN (2011) Cereal pollen sensitisation in pollen allergic patients: to treat or not to treat? *Eur Ann Allergy Clin Immunol* 43, 36–44.
- Darkó E, Ambrusa H, Stefanovits-Bányai E, Fodor J, Bakos F, and Barnabás B (2004) Aluminum toxicity, Al tolerance and oxidative stress in an Alsensitive wheat genotype and in Al-tolerant lines developed by in vitro microspore selection. *Plant Sci* 166, 583–91.
- da Silva ALS, Sperling P, Horst W, Franke S, Ott C, Becker D et al. (2006) A possible role of sphingolipids in the aluminum resistance of yeast and maize. J Plant Physiol 163, 26–38.
- Delhaize E, Gruber BD, and Ryan PR (2007) The roles of organic anion permeases in aluminum resistance and mineral nutrition. *FEBS Lett* **581**, 2255–62.
- Delhaize1 E, Ma JF, and Ryan PR (2012) Transcriptional regulation of aluminum tolerance genes. *Trends in Plant Sci* **17**, 341–8.
- Doncheva S, Amenós M, Poschenrieder C, and Barceló J (2005) Root cell patterning: a primary target for aluminum toxicity in maize. J Exp Bot 56, 1213–20.
- Doyle JJ and Luckow MA (2003) The rest of the iceberg. Legume diversity and evolution in a phylogenetic context. *Plant Physiol* 131, 900–10.
- Éva Darkó, Helga Ambrus, Éva Stefanovits-Bányai, József Fodor, Ferenc Bakos, and Beáta Barnabás (2004) Aluminium toxicity, Al tolerance and oxidative stress in an Al-sensitive wheat genotype and in Al-tolerant lines developed by in vitro microspore selection. *Plant Sci* 166, 583–91.
- Garzon T, Gunse B, Moreno AR, Tomos AD, Barcelo J, and Poschenrieder C (2011) Aluminium-induced alteration of ion homeostasis in root tip vacuoles of two maize varieties differing in Al tolerance. *Plant Sci* 180, 709–15.
- Gaume A, Mächler F, and Frossard E (2001) Aluminum resistance in two cultivars of *Zea mays* L., Root exudation of organic acids and influence of phosphorus nutrition. *Plant Oil* 234, 73–81.
- Godbold DL and Brunner I (2007) The platform for European root science, COST action E38: an introduction and overview. *Plant Biosyst* 141, 390–3.
- Graham PH and Vance CP (2003) Legumes: importance and constraints to greater use. *Plant Physiol* 131, 872–7.
- Gunsé B, Poschenrieder C, and Barcelo J (2000) The role of ethylene metabolism in the short-term responses to aluminium by roots of two maize cultivars different in Al-resistance. *Environ Exp Bot* 43, 73–81.
- Gunsé B, Garzón T, and Barceló J (2003) Study of aluminum toxicity by means of vital staining profiles in four cultivars of *Phaseolus vulgaris* L. *J Plant Physiol* 160, 1447–50.
- Guo TR, Yao PC, Zhang ZD, Wang JJ, and Wang M (2012) Devolvement of antioxidative defense system in rice growing seedlings exposed to Aluminum toxicity and Phosphorus deficiency. *Rice Sci* 19, 179–85.
- Hiradate S, Ma JF, and Matsumoto H (2007) Strategies of plants to adapt to mineral stresses in problem soils. In *Advances in Agronomy*. Donald, LS (ed.), pp. 65–132, Academic Press, New York, USA.
- Hirano Y and Brunner I (2006) Quantitative determination of callose in tree roots. J Plant Physiol 163, 1333–6.
- Horst WJ, Wang YX, and Eticha D (2010). The role of root apoplasm in aluminum induced inhibition of root elongation and aluminum resistance of plants: a review. Ann Bot 106, 185–97.
- Hungria M and Vargas MAT (2000) Environmental factors affecting N_2 fixation in grain legumes in the tropics, with an emphasis on Brazil. *Field Crops Res* **65**, 151–64.
- Ismail M (2005) Aluminum-phosphorus interactions on growth and some physiological traits of carrot and radish plants. *Acta Agron Hung* 53, 293–301.

- Kidd PS, Llugany M, Poschenrieder CH, Gunse B, and Barcelo J (2001) The role of root exudates in aluminium resistance and silicon-induced amelioration of aluminium toxicity in three varieties of maize (*Zea mays* L.). J Exp Bot 52, 1339–52.
- Kikui S, Sasaki T, Maekawa M, Mryao A, Hirochika H, Matsumoto H et al. (2005) Physiological and genetic analyses of aluminum tolerance in rice, focusing on root growth during germination. *J Inorg Biochem* 99, 1837– 44.
- Kikui S, Sasaki T, Osawa H, Matsumoto H, and Yamamoto Y (2007) Malate enhances recovery from aluminum-caused inhibition of root elongation in wheat. *Plant Soil* 290, 1–15.
- Klug B and Horst WJ (2010) Oxalate exudation into the root-tip water free space confers protection against aluminum toxicity and allows aluminum accumulation in the symplast in buckwheat (*Fagopyrum esculentum*). *New Phytol* 187, 380–91.
- Kochian LV (2004) How do crop plants tolerate acid soils? Mechanisms of aluminium tolerance and phosphorus efficiency. *Annu Rev Plant Biol* 55, 459–93.
- Kochian LV, Piñeros MA, and Hoekenga OA (2005) The physiology, genetics and molecular biology of plant aluminium resistance and toxicity. *Plant Soil* 274, 175–95.
- Kollmeier M, Felle HH, and Horst WJ (2000) Genotypical differences in aluminum resistance of maize are expressed in the distal part of the transition zone; Is reduced basipetal auxin flow involved in inhibition of root elongation by aluminum? *J Plant Physiol* **122**, 945–56.
- Kuo MC and Kao CH (2003) Aluminum effects on lipid peroxidation and antioxidative enzyme activated in rice leaves. *Biol Plant* 46, 149–52.
- León-Ruiz E, Alcázar P, Domínguez-Vilches E, and Galán C (2011) Study of Poaceae phenology in a Mediterranean climate. Which species contribute most to airborne pollen counts? *Aerobiologia* 27, 37–50.
- Li CS, Liu P, Xu GD, and Lin HJ (2006) Ameliorating effects of exogenous organic acids on aluminum toxicity in buckwheat seedlings. J Acta Agronomica Sinica 32, 532–9.
- Li CS, Liu P, Xu GD, He WB, and Zhu J (2004) Effect of acid-Al on the germination of soaked buckwheat seeds. *J Seed* 23, 9–11.
- Li XF, Ma JF, and Matsumoto H (2000) Pattern of Aluminum-induced secretion of organic acids differs between rye and wheat. *Plant Physiol* 123, 1537–43.
- Lilienfein J, Qualls RG, Uselman SM, and Bridgham SD (2003) Soil formation and organic matter accretion in a young andesitic chronosequence at Mt. Shasta, California. *Geoderma* 116, 249–64.
- Liu J, Magalhaes JV, Shaff J, and Kochian LV (2009) Aluminum activated citrate and malate transporters from the MATE and ALMT families function independently to confer Arabidopsis aluminum tolerance. *Plant* J 57, 389–99.
- Ma FJ, Ryan PR, and Delhaize E (2001) Aluminum tolerance in plants and the complexing role of organic acids. *Trends Plant Sci* **6**, 273–8.
- Ma JF and Furukawa J (2003) Recent progress in the research of external Al detoxification in higher plants: a mini review. J Inorg Biochem 97, 46– 51.
- Ma JF, She RF, Nagao S, and Tanimoto E (2004) Aluminium targets elongating cells by reducing cell wall extensibility in wheat roots. *Plant Cell Physiol* 45, 583–9.
- Maron LG, Piñeros MA, Guimaraes CT, Magalhaes JV, Pleiman JK, Mao CZ et al. (2010) Two functionally distinct members of the MATE (multidrug and toxic compound extrusion) family of transporters potentially underlie two major aluminum tolerance QTLs in maize. *Plant J* 61, 728– 40.
- Matsumoto H (2000) Cell biology of aluminum toxicity and tolerance in higher plants. *Int Rev Cytol* **200**, 1–46.
- Matsumoto H and Motoda H (2012) Aluminum toxicity recovery processes in root apices; possible association with oxidative stress. *Plant Sci* 186, 1–8.
- Meda AR and Furlani PR (2005) Tolerance to aluminum toxicity by tropical leguminous plants used as cover crops. *Braz Arch Biol Technol* 48, 309– 17.

Meriga B, Attitalla IH, Ramgopal M, Ediga A, and Kavikishor PB (2010)

Differential tolerance to Aluminum toxicity in rice cultivars: Involvement of antioxidative enzymes and possible role of Aluminum resistant locus. *Acad J Plant Sci* **3**, 53–63.

- Motoda H, Kano Y, Hiragami F, Kawamura K, and Matsumoto H (2010) Morphological changes in the apex of pea roots during and after recovery from aluminum treatment. *Plant Soil* **333**, 49–58.
- Motoda H, Kano Y, Hiragami F, Kawamura K, and Matsumoto H (2011) Changes in rupture formation and zonary region stained with Evans blue during the recovery process from aluminum toxicity in the pea root apex. *Plant Signal Behav* **6**, 98–100.
- Nagy NE, Dalen LS, Jones DL, Swensen B, Fossdal CG, and Eldhuset TD (2004) Cytological and enzymatic responses to aluminum stress in root tips of Norway spruce seedlings. *New Phytol* 163, 595–607.
- Nasr N, Carapetian J, Heidari R, Asri Rezaei S, Abbaspour N, and Darvishzadeh R (2011) Differential effects of aluminum on the seedling parameters of wheat. *African J Biotechnol* 10, 3345–53.
- Osawa H and Matsumoto H (2001) Possible involvement of protein phosphorylation in aluminium-responsive malate efflux from wheat root apex. *Plant Physiol* **126**, 411–20.
- Peixoto PHP, Cambraia J, Sant'Anna R, Mosquim PR, and Moreira MA (2001) Aluminum effects on fatty acid composition and lipid peroxidation of a purified plasma membrane fraction of root apices of two sorghum cultivars. J Plant Nutr 24, 1061–70.
- Piñeros MA, Magalhaes JV, Alves VMC, and Kochian LV (2005) Aluminum resistance in maize cannot be solely explained by root organic acid exudation. A comparative physiological study. *Plant Physiol* 137, 231– 41.
- Poozesh V, Cruz P, Choler P, and Bertoni G (2007) Relationship between the Al resistance of grasses and their adaptation to an unfertile habitat. *Ann Bot* **99**, 947–54.
- Poschenrieder C, Tolra' RP, and Barcelo J (2005) A role for cyclic hydroxamates in aluminium resistance in maize? J Inorg Biochem 99, 1830–6.
- Poschenrieder C, Gunsé B, Corrales I, and Barceló J (2008) A glance into aluminum toxicity and resistance in plants. *Sci Total Environ* 400, 356– 68.
- Purcino AAC, Alves VMC, Parentoni SN, Belele CL, and Loguercio LL (2003) Aluminum effects on nitrogen uptake and nitrogen assimilating enzymes in maize genotypes with contrasting tolerance to aluminum toxicity. J Plant Nutr 26, 31–61.
- Rangel AF, Mobin M, Rao IM, and Horst WJ (2005) Proton toxicity interferes with the screening of common bean (*Phaseolus vulgaris* L.) genotypes for aluminium resistance in nutrient solution. J Plant Nutr Soil Sci 168, 607–16.
- Rangel AF, Rao IM, and Horst WJ (2009) Intracellular distribution and binding state of aluminum in root apices of two common bean (*Phaseolus vulgaris*) genotypes in relation to Al toxicity. *Physiol Plant* 135, 162–73.
- Ruf M and Brunner I (2003) Vitality of tree fine roots: reevaluation of the tetrazolium test. *Tree Physiol* 23, 257–63.
- Ryan PR, Delhaize E, and Jones DL (2001) Function and mechanism of organic anion exudation from plant roots. *Ann Rev Plant Physiol Plant Molec Biol* 52, 527–60.
- Ryan PR, Tyerman SD, Sasaki T, Furuichi T, Yamamoto Y, Zhang WH et al. (2011) The identification of aluminium-resistance genes provides opportunities for enhancing crop production on acid soils. *J Exp Bot* 62, 9–20.
- Sarker BC and Karmoker JL (2011) Effects of phosphorus deficiency on accumulation of biochemical compounds in lentil (*Lensculinaris* medik.). *Bangladesh J Bot* **40**, 23–7.
- Sharma P and Dubey RS (2007) Involvement of oxidative stress and role of antioxidative defense system in growing rice seedlings exposed to toxic concentrations of aluminum. *Plant Cell Rep* **26**, 2027–38.
- Silva S (2012) Aluminum toxicity targets in plants. J Bot 2012, 219-62.
- Singh NB, Yadav K, and Amist N (2011) Phytotoxic effects of Aluminum on growth and metabolism of *Pisum sativum L. Int J Innov Biol Chem Sci*

2, 10–21.

- Smith E, Naik D, and Cumming JR (2011) Genotypic variation in aluminum resistance, cellular aluminum fractions, callose and pectin formation and organic acid accumulation in roots of Populus hybrids. *Environ Expl Bot* 72, 182–93.
- Sun P, Tian QY, Chen J, and Zhang WH (2010) Aluminum-induced inhibition of root elongation in Arabidopsis is mediated by Ethylene and Auxin. J Exp Bot 61, 346–56.
- Sun P, Tian QY, Zhao MG, Dai XY, Huang JH and Zhang WH (2007) Aluminum-induced ethylene production is associated with inhibition of root elongation in *Lotus japonicus* L. *Plant Cell Physiol* 48, 1229–35.
- Teraoka T, Kaneko M, Mori S, and Yoshimura E (2002) Aluminum rapidly inhibits cellulose synthesis in roots of barley and wheat seedlings. J Plant Physiol 159, 17–23.
- Tewari RK, Kumar P, Tewari N, Srivastava S, and Sharma PN (2004) Macronutrient deficiencies and differential antioxidant responsesinfluence on the activity and expression of superoxide dismutase in maize. *Plant Sci* 66, 687–94.
- Tolra R, Barcelo J, and Poschenrieder C (2009) Constitutive and aluminiuminduced patterns of phenolic compounds in two maize varieties differing in aluminium tolerance. *J Inorg Biochem* **103**, 1486–90.
- Tolra R, Poschenrieder C, Lupi B, and Barcelo J (2005) Aluminium-induced changes in the profiles of both organic acids and phenolic substances underlie Al tolerance in *Rumex acetosa* L. *Environ Exp Bot* 54, 231–8.
- Van Wyk BE (2005) Food plants of the world. Identification, culinary uses and nutritional value. Briza Publications, Pretoria, South Africa.
- Vanguelova EI, Hirano Y, Eldhuset TD, Sas-Paszt L, Bakker MR, Püttsepp Ü et al. (2007) Tree fine root Ca/AI molar ratioindicator of AI and acidity

stress. Plant Biosyst 141, 460-80.

- Wang JW and Kao CH (2004) Reduction of aluminum-inhibited root growth of rice seedlings with supplemental calcium, magnesium and organic acids. Crop Environ Bioinform 1, 191–8.
- Wang P, BI S, and Han W (2006) Aluminum tolerance of two wheat cultivars (Brevor and Atlas66) in relation to their rhizosphere pH and organic acids exuded from roots. J Agric Food Chem 54, 10033–9.
- Xiao XX, Yang ZW, Zhong R, and Chen L (2003) Effect of aluminum stress on cell ultrastructure of leaf and root of longan. *Scientia Silvae Sinicae* 23, 58–63.
- Ying XF, Liu P, Xu GD, Lu QD, and Zhu SL (2005) Screening of soybean genotypes with tolerance to aluminum toxicity and study of the screening indices. *Chin J Oil Crop Sci* 27, 46–51.
- Yu HN, Liu P, Wang ZY, Chen WR, and Xu GD (2011) The effect of aluminum treatments on the root growth and cell ultrastructure of two soybean genotypes. *Crop Protection* 30, 323–8.
- Zakir H, Koyama H, and Hara T (2003) Growth and cell wall properties of two wheat cultivars differing in their sensitivity to aluminum stress. J Plant Physiol 163, 39–47.
- Zhang WH, Ryan PR, and Tyerman SD (2001) Malate-permeable channels and cation channels activated by aluminium in the apical cells of wheat roots. *Plant Physiol* **125**, 1459–72.
- Zheng SJ and Yang JL (2005) Target sites of aluminum Phytotoxicity. *Biol Plant* **49**, 321–31.
- Zhou S, Sauvé R, and Thannhauser TW (2009) Proteome changes induced by aluminum stress in tomato roots. *J Exp Bot* **57** 4201–13.
- Zobel RW and Kinraide TB (2007) Fine root diameters can change in response to nutrient concentrations. *Plant Soil* **297**, 243–54.