



In vitro Antiurolithiatic Potential in *Asparagus falcatus* L., A Folklore Medicinal Plant in Sri Lanka

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Abstract

Urolithiasis is the condition where urinary calculi are formed in the urinary tract and is identified as one of the most prevalent diseases throughout the globe. However, the treatment approaches employed in modern medicine are not capable of preventing the recurrence of urinary calculi formation. In this respect, the plant species used in traditional systems of medicine to treat and/or prevent urinary calculi could be explored in order to develop novel antiurolithiatic agents. Therefore, the present study aims at evaluating the antiurolithiatic potential in *Asparagus falcatus* L., a plant widely employed in indigenous medicine in Sri Lanka to dissolve urinary calculi and to treat other urinary disease conditions. Thus, herein, the antiurolithiatic potential of methanolic extract of the root of *A. falcatus* was studied under *in vitro* conditions using crystal nucleation and aggregation assays. Furthermore, the dissolution assay was employed to determine the ability of the plant extract in dissolving surgically removed urinary calculi samples from patients and the results were expressed as percentage weight loss of calculi sample. The results of the *in vitro* assays revealed that the above extract is capable of preventing crystal nucleation by 13.26% and aggregation by 36.5% at the concentration of 2000 µg/mL. Its efficacy was comparable with a marketed polyherbal combination, cystone which displayed 17.4% and 44.27% inhibition of nucleation and aggregation respectively at the above concentration. In the dissolution assay, the plant extract reduced the weight of the calculi by 5.19% in comparison to the weight loss of 4.28% in the positive control cystone. Thus, our study indicated that the roots of *A. falcatus* was a potent and promising antiurolithiatic agent while rationalizing its use in traditional medicine.

Keywords: Antiurolithiatic, *Asparagus falcatus*, herbal, urinary calculi

Introduction

The term "urolithiasis" is used to describe the deposition or formation of urinary calculi in any part of the urinary system [1]. It is a common health problem with an increasing prevalence of up to 20% throughout the globe [2]. The development of the calculi is related to the decrease of urine volume or the increase of excretion of calculi-forming solute materials such as calcium, oxalate, urate, cystine, xanthine, and phosphate where calcium oxalate has been identified as the most predominant component of urinary calculi [2]. A number of sequential events involved in the calculi formation, namely; urinary saturation, supersaturation, nucleation, crystal growth, aggregation of crystals, crystal retention, and, finally, the calculus formation [3].

The surgical removal of urinary calculi is gradually replacing with the modern techniques like extracorporeal lithotripsy, ureteroscopy and local calculus disruption using a high power laser, however, none of these therapeutic approaches could avoid the recurrence of renal calculi, in which the recurrence rate is estimated nearly 50% [4]. Since the pathogenesis of urolithiasis is attributed by multiple factors, still a satisfactory drug is not available for the treatment as well as to prevent the recurrence of the calculi [5]. Moreover, hypocalciuric agents employed as treatment modalities have several adverse effects which necessitate the development of novel strategies for the prevention and treatment of urinary calculi [6]. In view of the above, the medicinal plants employed in indigenous medicine to treat urinary diseases could be potential sources of novel antiurolithiatic agents.

The genus *Asparagus* (Family: Asparagaceae) comprises of about 100 species that are widespread in the old World [7]. Several species of *Asparagus* are either grown naturally or cultivated in Sri Lanka, out of which *A. falcatus* L., ("Hathawariya") and *A. racemosus* Willd., ("Heen Hathawariya") are widely employed in indigenous medicine in the country as a remedy for chronic nephritis, as an antilithic for urinary gravel and calculi as well as a diuretic agent [8, 9]. Although both these

species are considered to be equally important as herbal remedies, most of the research studies were focused on *A. racemosus*, but not on *A. falcatus*. For example, the antiurolithiatic activity in ethanolic extract of *A. racemosus* was scientifically validated by both *in vitro* and *in vivo* studies [10,11] while various secondary metabolites such as shatvarin, immunoside, asparagine A, asparagine B etc. have been isolated from this plant [12]. However, the scientific evidence are scarce to rationalize the extensive utility of *A. falcatus* as an antiurolithiatic agent in indigenous medicine. Thus the present study was undertaken to evaluate the antiurolithiatic activity of roots of *A. falcatus*, a plant that has been used from the time immemorial by traditional healers in Sri Lanka for the treatment of urinary calculi.

Materials and Methods

Preparation of plant extract

Roots of *A. falcatus* were collected from Gampaha district in Western Province of Sri Lanka in 2017 and the plant material was authenticated from the National Herbarium, Royal Botanical Gardens, Peradeniya, Sri Lanka. A voucher specimen (WP2017-NO_05) was deposited at the Department of Biochemistry, Faculty of Medicine, University of Ruhuna, Sri Lanka for future references. The shade dried plant materials (20 g) were extracted in methanol (Sigma- Aldrich) for two days. Thereafter, the liquid aliquot obtained by filtration was evaporated using a rotary evaporator (HAHNAPOR, HS-2005V) into complete dryness and six predetermined test concentrations (2000, 1000, 500, 250, 125 and 62.5 µg/mL) were prepared from this crude extract.

Determination of antiurolithiatic activity

Nucleation assay

The spectroscopic method of Hennequin et al [13] was employed with slight modifications as described by Napagoda et al. [14], to

determine the inhibitory activity of the extracts on the nucleation of CaC_2O_4 crystals. Briefly, CaCl_2 (4 mmol/L) and $\text{Na}_2\text{C}_2\text{O}_4$ (50 mmol/L) solutions were added to artificial urine in the presence of plant extract at different concentrations in order to initiate the crystallization. The nucleation was determined by the appearance of crystals that reached a critical size and became optically detectable either in the presence of the extract or without the extract (negative control). The absorbance was measured at 620 nm, and the percentage inhibition was calculated as $[(C - S)/C] \times 100$, where, C is the turbidity without plant extract, S is the turbidity with plant extract.

Cystone, a marketed polyherbal formulation that could prevent the formation of lithogenic substances and disintegrate urinary calculi [15] was used as the positive control.

Aggregation assay

The method of Hess et al. [16], was used with slight modifications as described by Napagoda et al. [14] in the determination of the rate of aggregation of the CaC_2O_4 crystals. In brief, CaC_2O_4 crystals were prepared by mixing CaCl_2 and $\text{Na}_2\text{C}_2\text{O}_4$ (50 mmol/L each) which were equilibrated in a bath for 1 h at 60 °C. Thereafter, the solutions were cooled to 37°C and then evaporated. The CaC_2O_4 crystals were dissolved with Tris (0.05 mol/L) and NaCl (0.15 mol/L) at pH 6.5 to a final concentration of 1 mg/mL. The reaction mixture was treated with the plant extract and the absorbance at 620 nm was recorded at 30, 60, 90, 180 and 360 min. The percentage inhibition was calculated as $[(C - S)/C] \times 100$, where, C is the turbidity without plant extract, S is the turbidity with plant extract. Cystone was used as the positive control.

Dissolution of urinary calculi

The method as described by Napagoda et al. [14], was used to determine the dissolution of urinary calculi. For this assay, the urinary calculi were obtained from three male patients in the age group of 30-50 years who were admitted to the Teaching Hospital, Karapitiya, Sri Lanka and underwent surgical treatment. Only the patients with a prior history of urinary calculi were selected to collect the calculi samples.

The collected calculi were crushed to homogenize the sample. A sample of 60 mg from this crushed materials was put into each of the test tubes, into which different concentrations of extracts were added. These were allowed 24, 48, and 72 hours for dissolution at 37 ± 0.5 °C and the weight of the calculi were obtained at each of the above time intervals. After the dissolution period, the reduction in weight and percentage weight loss were calculated using the following formula.

$\% \text{ Weight loss} = [(W_i - W_f)/W_i] \times 100$ where W_i is the initial weight of the calculi and W_f is the final weight of the calculi.

Cystone was used as the positive control while distilled water was used as the negative control.

Statistical analysis

All experiments were conducted in duplicates and all data were presented as mean \pm standard deviation (SD). Statistical evaluation of the data was performed by one-way ANOVA using Minitab version 14. A p value < 0.05 was considered significant.

Qualitative analysis of the urinary calculi obtained from patients

The powdered urinary calculi were tested for the presence of inorganic and organic constituents by standard qualitative tests described by Hodgkinson [17] with slight modifications. A brief outline of the protocol employed is illustrated in Figure 1 and described in detail

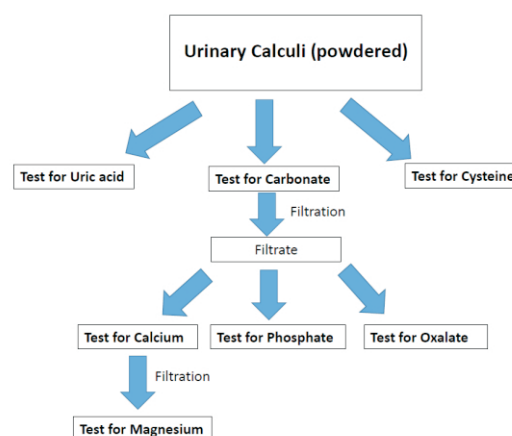


Figure 1: Testing of urinary calculi for the presence of organic and inorganic constituents

Test for uric acid

KOH was added to the powdered calculi sample and the filtrate was collected. Follin's reagent and NaCN were added to this filtrate and observed for the development of a blue colored solution.

Test for cysteine

The powdered sample was placed on a white tile into which NaCN and freshly prepared sodium nitroprusside solutions were added and was observed for the development of magenta color.

Test for carbonate

The powdered urinary calculi sample was treated with HNO_3 acid and observed for effervesce as an indication of the presence of carbonate. Thereafter the resulted filtrate was divided into three parts and each portion was subjected to the following tests.

Test for calcium and magnesium

A few drops of $(\text{NH}_4)_2\text{C}_2\text{O}_4$ was added to the one portion of the filtrate and its pH was adjusted to pH 5. The reaction mixture was observed for the presence of a precipitate.

Subsequently, this precipitate was completely filtered off and a few drops of K_3PO_4 and ammonia were added to the filtrate until it became alkaline and thereafter observed for the development of crystalline precipitate.

Test for phosphate

The second portion of the filtrate was treated with ammonium molybdate and allowed to stand at room temperature. This reaction mixture was observed for the development of yellow colored precipitate which turns blue upon the addition of reducing agents.

Test for oxalate

The third portion of the filtrate was treated with a few drops of CaCl_2 and the pH was adjusted to pH 5. The reaction mixture was observed for the formation of a precipitate. Thereafter, the precipitate was filtered off and the filter paper was washed with hot 0.25 M H_2SO_4 . A few drops of diluted KMnO_4 were added to this washed solution and was observed for a color change.

Ethical consideration

Ethical approval was obtained from the Ethical Review Committee, Faculty of Allied Health Sciences, University of Ruhuna, Sri Lanka.

Results and Discussion

Nucleation assay

The pathogenesis of urinary calculi formation is a multifaceted process that involves urine saturation and super-saturation, crystal nucleation, aggregation, the retention of crystals by the urothelium, and the continued growth of the stone on the retained crystals [18]. The formation of the nucleus from supersaturated urine is the first step in the formation of urinary calculi [19], thus the classical model for the study of oxalate crystallization was employed here to study the formation and growth of the calcium oxalate monohydrate crystals from artificial urine.

The results indicated that the inhibition of nucleation was concentration dependent (Figure 2) and the maximum inhibition was observed as 13.26% for the plant extract and as 17.4% for the positive control cystone at the concentration of 2000 $\mu\text{g/mL}$ (Figure 3). Interestingly, there was no significant difference in inhibitory potency of the test extract and the positive control ($p = 0.762$).

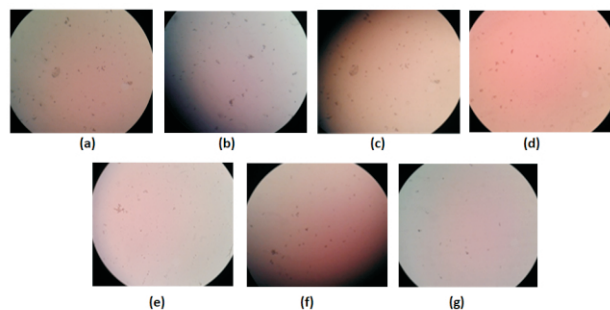


Figure 2: Microscopic evaluation of different concentrations of *A. falcatus* extract on CaC_2O_4 nucleation (40 \times magnification)

(a) Negative control (without the extract) (b) 62.5 $\mu\text{g/mL}$ (c) 125 $\mu\text{g/mL}$ (d) 250 $\mu\text{g/mL}$ (e) 500 $\mu\text{g/mL}$ (f) 1000 $\mu\text{g/mL}$ (g) 2000 $\mu\text{g/mL}$.

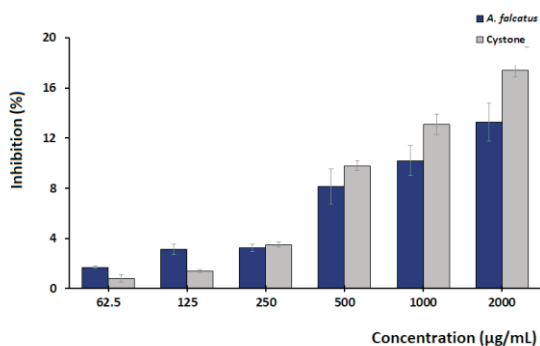


Figure 3: Effect of *A. falcatus* extract and cystone on CaC_2O_4 nucleation at different concentrations

Aggregation assay

The small hard mass of urinary crystals in solution sticks together to form larger calculi through a process known as

aggregation/agglomeration which is considered to be the most critical step in calculi formation [19]. Therefore, the inhibitory potential of *A. falcatus* extract on crystal aggregation was studied (Figure 4) and compared with the commercial polyherbal remedy, cystone. The highest inhibition of crystal aggregation was observed as 36.5% for the plant extract and 44.27% for cystone at 2000 $\mu\text{g/mL}$ (Figure 5). The statistical analysis revealed that there was no significant difference between the inhibitory potency of the plant extract and the positive control ($p = 0.518$), indicating that both were equally effective. However, the inhibition of aggregation was not found to be time dependent.

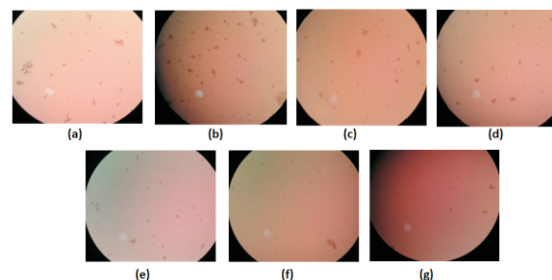


Figure 4: Microscopic evaluation of different concentration of *A. falcatus* extract on CaC_2O_4 crystal aggregation (40 \times magnification). (a) Negative control (without the extract) (b) 62.5 $\mu\text{g/mL}$ (c) 125 $\mu\text{g/mL}$ (d) 250 $\mu\text{g/mL}$ (e) 500 $\mu\text{g/mL}$ (f) 1000 $\mu\text{g/mL}$ (g) 2000 $\mu\text{g/mL}$.

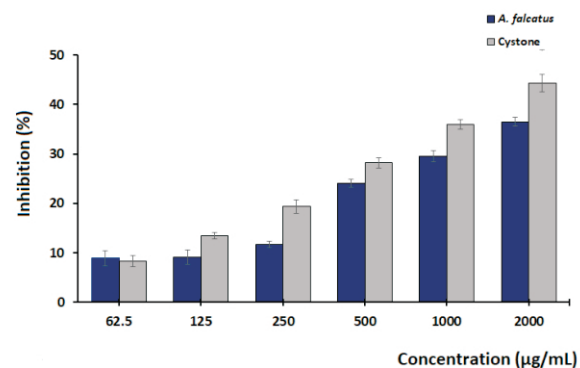


Figure 5: Effect of *A. falcatus* extract and cystone on CaC_2O_4 aggregation at different concentrations

Dissolution of urinary calculi

The efficacy in the dissolution of urinary calculi was expressed as percentage weight change at different time intervals. It was found to be time dependent as the highest mean percentage weight loss of urinary calculi was observed after 72 hours. However, the percentage weight loss was not found to be concentration dependent as the highest value was observed at 500 $\mu\text{g/mL}$ concentration for both the plant extract and the positive control, cystone. Interestingly, the plant extract was found to be more effective than the positive control with a weight change of 5.19% in comparison to the weight change of 4.28% in the positive control (Figure 6). The statistical analysis revealed that a significance difference does exist ($p=0.04$) between these two treatments.

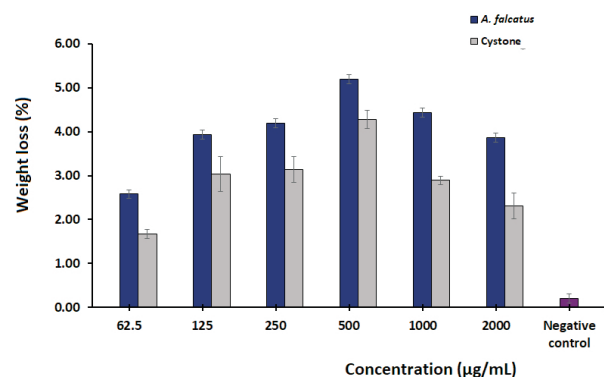


Figure 6: Effect of *A. falcatus* extract and cystone on dissolution of urinary calculi at different concentrations after incubation for 72 hours

Qualitative analysis of the urinary calculi

The results of the qualitative analysis indicated the presence of carbonate, calcium, magnesium, phosphate, oxalate, uric acid and cysteine in the urinary calculi used in the dissolution assay.

Due to the unavailability of effective drugs to be used in the clinical therapy to prevent the recurrence of urinary calculi, as of recent time, the traditional medicinal plants have drawn the attention as potential sources to be explored as alternative antiuro lithiatic agents. In this respect, we assumed that the present study could provide valuable insight into the field of herbal antiuro lithiatic agents while validating the ethnobotanical significance of *A. falcatus* in the Sri Lankan folklore medicine.

In this work, the effect of *A. falcatus* extract on the calcium oxalate crystal nucleation and aggregation as well as the dissolution of urinary calculi composed of calcium, magnesium, phosphate, carbonate, oxalate, uric acid and cysteine were evaluated. These *in vitro* investigations revealed that *A. falcatus* is highly effective as an antiuro lithiatic agent by inhibiting the calcium oxalate crystal nucleation and aggregation in a concentration-dependent manner, similar to the positive control, cystone. Moreover, the plant extract is potent in the dissolution of urinary calculi and this potency surpassed that of cystone. The standard drug cystone has been identified as an effective formulation for the treatment of urolithiasis where the antilithiatic effect has been attributed to complex spectrum of actions including antimicrobial, diuretic, antispasmodic, litholytic, anti-inflammatory and anticalcifying activities of its ingredients [20]. However, under *in vitro* conditions, it is possible to hypothesize that the litholytic and anticalcifying potencies of cystone would make significant contributions to its antilithiatic effect, as other bioactivities are imposed mainly under *in vivo* situations. Similarly, the methanolic extract of *A. falcatus* may also exert several modes of actions that could be correlated with its antilithiatic effect, in addition to its anticalcifying and litholytic potencies observed in this *in vitro* study. Thus, more detailed *in vivo* experiments are required to elaborate the other possible underlying mechanisms involved with the antilithiatic activity of this extract. Nevertheless, the observed potencies in the extract are advantageous for the development of a product which is capable of preventing urinary calculi formation as well as to be used as an effective alternative therapy for urinary calculi. The observed bioactivities could be attributable to the presence of antiuro lithiatic phytochemicals in this extract, thus phytochemical analysis is warranted to identify the bioactive secondary metabolites while the scientific validation of its efficacy and safety could be achieved by detailed *in vivo* assays and cytotoxicity studies.

Conclusion

Our investigations indicated that under *in vitro* conditions, methanolic extract of the root of *A. falcatus* has a potent antiuro lithiatic activity due

to its inhibitory effects on the crystal nucleation, aggregation as well as dissolution of urinary calculi and these observations rationalize the folklore claims of this plant as an effective antiuro lithiatic agent.

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Conflict of interest

The authors declare that they have no competing interest.

References

- [1] Agarwal K, Varma R. *Ocimum gratissimum* L.: A medicinal plant with promising antiuro lithiatic activity. *Int J Pharm Sci Drug Res* 2014; 6(1):78-81.
- [2] Yachi L, Bennis S, Aliat Z, Cheikh A, Idrissi MOB, Draoui M, Bouatia M. *In vitro* litholytic activity of some medicinal plants on urinary stones. *Afr. J. Urol* 2018; 24(3):197-201.
- [3] García-Perdomo HA, Solarte PB, España PP. Pathophysiology associated with forming urinary stones. *Urol Colomb* 2016; 25(2):118-25.
- [4] Saha S, Verma RJ. Inhibition of calcium oxalate crystallization *in vitro* by an extract of *Bergenia ciliata*. *Arab J Urol* 2013; 11:187-92.
- [5] Khan A, Khan SR, Gilani AH. Studies on the *in vitro* and *in vivo* antiuro lithiatic activity of *Holarrhena antidysenterica*. *Urol Res* 2012; 40(6):671-81.
- [6] Butterweck V, Khan S. Herbal Medicines in the Management of Urolithiasis: Alternative or Complementary? *Planta Med* 2009; 75(10):1095-1103.
- [7] Dinan, L, Savchenko T, Whiting. P. Phytoecdysteroids in the genus *Asparagus* (Asparagaceae). *Phytochemistry* 2001; 56(6):569-76.
- [8] Dassanayake MD, Clayton WD. A Revised Handbook of the Flora of Ceylon -Volume 14A A Balkema Publishers, Rotterdam, Netherlands, 2000: 94-98.
- [9] Jayaweera DMA. Medicinal plants (Indigenous and exotic) used in Ceylon- Part III, National Science Council Sri Lanka, 1982: 262-65.
- [10] Alok S, Sabharwal M, Mishra SB, Singh P, Singh M. *In vitro* evaluation on antiuro lithiatic activity of roots of *Asparagus racemosus* Willd. *Fl. & Fauna (Jhansi)* 2009; 15(1):163-66.
- [11] Jagannath N, Chikkannasetty SS, Govindadas D, Devasankaraiah G. Study of antiuro lithiatic activity of *Asparagus racemosus* on albino rats. *Indian J Pharmacol* 2012; 44(5):576-79.
- [12] Alok S, Jain SK, Verma A, Kumar M, Mahor A, Sabharwal M. Plant profile, phytochemistry and pharmacology of *Asparagus racemosus* (Shatavari): A review. *Asian Pac J Trop Dis* 2013; 3(3):242-51.
- [13] Hennequin C, Lalanne V, Daudon M, Lacour B, Druke T. A new approach to studying inhibitors of calcium oxalate crystal growth. *Urol Res* 1993; 21:101-08.
- [14] Napagoda M, Madhumadhavi N, Vimukthi K. Evaluation of antiuro lithiatic potential in *Crataeva religiosa*: An *in vitro* study. *Prayog Ras.* 2019; 3(2):1-4.
- [15] Devkar RA, Chaudhary S, Adepu S, Xavier SK, Chandrashekar KS, Setty MM. Evaluation of antiuro lithiatic and antioxidant potential of *Lepidagathis prostrata*: A Pashanbhed plant. *Pharm Biol* 2016; 54(7):1237-45.
- [16] Hess B, Nakagawa Y, Coe FL. Inhibition of calcium oxalate monohydrate crystal aggregation by urine proteins. *Am J Physiol* 1989; 257:99-106.
- [17] Hodgkinson A. A combined qualitative and quantitative procedure for the chemical analysis of urinary calculi. *J Clin Pathol* 1971; 24(2):147-51.
- [18] Mandel N. Mechanism of stone formation. *Semin Nephrol* 1996; 16(5):364-74.
- [19] Alelign T, Petros B. Kidney stone disease: An update on current concepts. *Adv Urol* 2018; 3068365. doi:10.1155/2018/3068365
- [20] Tiwari P, Kothiyal P, Ratan P. Antiuro lithiatic effect of some polyherbal formulations used in experimentally induced urolithiasis: A review. *Int Res J Pharm* 2017; 8(5):14-22.