Short Communication Studies on strain specificity of Frankia in Alnus nepalensis

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ABSTRACT

To identify an effective and compatible Frankia strain for Alnus nepalensis, experiments were carried out with four different strains. Alnus was infected by all the four Frankia strains as was evidenced by the higher growth parameters over the uninoculated control. The strain AVC - II performed better in terms of infectivity and productivity of Alnus when compared with other strains tested. The study clearly indicated the possibility of exploiting the host - endophyte specificity in Alnus for higher productivity through effective symbiosis.

Key words: *Alnus nepalensis*, *Frankia*, inoculation.

Nepalese alder (Alnus nepalensis) belonging to the family Betulaceae is one of 15 genera of trees that fix nitrogen but are not in legume family. Alnus occurs throughout the Himalaya hills at 500 - 3000 m elevation from Pakistan through Nepal, Bhutan and Burma to southwest China. It is found naturally in subtropical mountains with an average rainfall of 500 - 2500 mm and a 4 - 8 month dry season. Alnus forms a symbiosis with nitrogen fixing actinomycetes of the genus Frankia. Frankia are known to infect some 165 plant species distributed among eight orders of plants (Bond 1983). Frankia-Alnus symbiosis increase the nitrogen content of soil by about 61.5 to 157 kg N ha⁻¹ year⁻¹ which indicates the importance of this association in the overall nitrogen economy of the soil (Subba Rao 1989).

Low nitrogen fixing Frankia strains can also infect the host, which suggests the need for inoculation of the Alnus roots with right endophyte before field planting. Efficient Frankia strains must be able to compete with wild strains to nodulate the host plant, resulting in a highly efficient association in the field. The right host-strain specificity should be assessed to find out the best combination for cultural use. Hence the experiments were conducted to identify an efficient Frankia strain which could establish good nodulation for higher productivity in Alnus nepalensis.

The experiments consisted of three *Frankia* strains and a nodule suspension of *Alnus nepalensis* besides an uninoculated control. The nodule suspension was prepared using active healthy nodules collected from natural stands. They were packed in plastic bags on ice and stored under - 10°C. Nodules were washed with sterile water and surface

sterilized with 30 % hydrogen peroxide for 5 minutes under aseptic conditions. The suspension was prepared by crushing nodules using mortar and pestle (30 g nodule: 100 ml sterile distilled water). The suspension thus obtained was used for inoculation.

The Frankia strains viz., Frankia 53024, Frankia H - 43 and Frankia AVC - II were obtained as pure cultures in the liquid broth from the Department of Botany, University of Glasgow, Glasgow, United Kingdom. Aseptically grown, one year old Alnus seedlings were dipped in the pure cultures of Frankia nodule suspension for 30 minutes and planted in the field at Horticultural Research Station farm, Udhagamandalam, The Nilgiri Hills. The experiments were laid out in a randomized block design with five treatments and four replications. Six months after inoculation, the effect of each treatment was evaluated for plant growth characters viz., root length, shoot length, plant biomass and total chlorophyll content. Infectivity of each strain was examined in terms of nodule number, size, dry weight and nodule nitrogenase activity by acetylene ethylene assay (Hardy et al. 1968).

Results indicated that *Alnus* was infected by all the strains tested, as was evidenced by the higher growth parameters when compared with the uninoculated control (Table 1). Among the inoculated treatments, higher productivity was observed in *Alnus* plants inoculated with *Frankia* AVC II, recording higher shoot length, root length, dry matter production and chlorophyll content followed by *Frankia* H 43 and *Frankia* 53024 strains. Compared with the uninoculated control, all

Table 1. Effect of inoculation of Alnus nepalensis with Frankia strains on plant growth and chlorophyll content.

Treatment	Shoot length (cm)	Root length (cm)	Total chlorophyll (mg g 'FW)	Plant dry weight (g)
Nodule suspension of Alnus nepalensis	38.00	13.56	2.40	4.62
2.Frankia 53024	49.65	17.56	2.65	5.92
3. Frankia H - 43	51.08	19.00	2.81	6.12
4. Frankia AVC - II	64.00	25 00	3.96	8.71
5 Control	23.00	11.50	2.18	3.10
LSD (0.05%)	4.004	1.033	0.306	0.366

FW Fresh weight

Table 2. Infectivity of Frankia strains on Alnus nepalensis.

Treatment	Nodule number	Nodule diameter (cm)	Nodule dry weight (mg)	Nodule nitrogenase activity (u mol C, H, g ' hr ')
Nodule suspension of Alnus nepalensis	6.14	0.34	2.60	0.34
2. Frankia 53024	8.62	0.46	3.21	1.59
3. Frankia H - 43	9.81	0 52	3.62	1.62
4. Frankia AVC - II	14.62	0.74	4.50	2.21
5 Control	3.38	0.21	0.51	0.21
LSD (0.05%)	0.472	0.909	0.344	0.237

treatments with *Frankia* inoculations helped in higher biomass built-up in *Alnus* (Simon *et al.* 1985). The infectivity assay (Table 2) also showed a similar trend: higher nodule number and nitrogenase activity when inoculated with *Frankia* AVC - II strain. The higher performance of the *Frankia* pure culture might be attributed to the high quality inoculum, high concentration of infective cells and long term viability conferred by spores (Van Dijk 1978).

The other two Frankia strains viz., Frankia 53024 and Frankia H 43 were similar in their performance, followed by the nodule suspension. This kind of infectivity variation among the Frankia strains confirmed the heterogenic infectivity of Frankia (Wheeler et al. 1991). The poor performance of these two Frankia strains compared to Frankia AVC - II may possibly be due to the lower proportion of high infective Frankia cells (Van Dijk and Slumier Stolk 1990) and possession of partial compatible Frankia strains (Torrey 1990). Nodules obtained under field conditions may contain more than one strain (Nesme et al., 1985) and this might be the reason for the lower infectivity of the nodule suspension.

The results showed the infectivity variation among the *Frankia* strains tested. However, all the strains ultimately helped in higher biomass built-up, nodule number and nitrogenase activity of *Alnus*. Therefore the results suggest the necessity for selecting an optimal combination of a host genotypeendophyte strain for increased productivity. *Frankia* strain AVC - II was found to be the best symbiont for *Alnus nepalensis* in this experiment.

REFERENCES

- Bond G 1983 Taxonomy and distribution of nonlegume nitrogen fixing systems. In: Biological Nitrogen Fixation in Forest Ecosystems. Gordon J C and Wheeler C T (Eds.), Nijhoff Publishers, The Hague. pp. 55 - 87.
- Hardy RWF, Holsten RD, Jackson EK and Burns RC 1968 The acetylene ethylene assay for N₂ fixation: Laboratory and field evaluation. Pl. Physiol. 43: 1185-1207.
- Nesme X, Normand P, Tremblay FM and Lalonde M 1985 Nodulation speed of *Frankia* species on *Alnus glutinosa*, *Alnus crispa* and *Myrica gale*. Can. J. Bot. 63: 1292-1295.
- Simon L S Cote A Stein and Lalonde M 1985 Performance of *in vitro* propagated *Alnus* glutinosa (L) Gaertner. clones inoculated with *Frankia*. Plant Soil. 87: 125 - 134.
- Subba Rao N S 1989 Nitrogen fixation by nodulated plants other than legumes. In: Soil Microorganisms and Plant Growth. Oxford and IBH PublishingCo., New Delhi. pp.184-191.
- Torrey JG 1990 Cross inoculation groups within *Frankia* host endosymbiont association. In: The Biology of *Frankia* and Actinorhizal Plants. Schwintzer CR and Tjepkema JD (Eds.), Academic Press, New York. pp. 83-106.
- Van Dijk C 1978 Spore formation and endophyte diversity in root nodules of *Alnus glutinosa* (L) Gaertner. New Phytol. 81: 601-615.
- Van Dijk C and Slumier Stolk A 1990 An ineffective strain type of *Frankia* in the soil of natural stands of *Alnus glutinosa* (L) Gaertner. Plant Soil. 127: 107-121.
- Wheeler CT, Miller IM, Narayanan R and Purushothaman D 1991 Soil micro organisms in agroforestry systems. In: Biophysical Research for Asian Agroforestry. Avery ME, Cannell MGR and Ong CK (Eds.), pp.143-166.