Fungi associated with maize seed discolouration and abnormalities in South-western Nigeria

B.F.Owolade', B. Fawole² and Y. O. K. Osikanlu¹

¹Institute of Agricultural Research and Training, Moor Plantation, Ibadan, Nigeria.

Accepted 08 May 2000

ABSTRACT

Three seed samples of maize from Ibadan in South-western Nigeria showing different forms of discolouration and abnormalities were screened for associated fungi. Fusarium moniliforme and Aspergillus flavus were isolated from all the categories of seed tested. The percentage incidence of F. moniliforme was significantly higher on seeds which showed white streaks, purple/pink discolouration, discoloured germ end and wrinkling, than that of any other fungus. Cephalosporium acremonium and Nigrospora oryzae were also associated with seeds that showed white streaks. Fusarium graminearum was associated with purple/pink discolouration while Drechslera maydis, Fusarium semitectum, Curvularia lunata and Colletotrichum graminicola were observed on seeds with brown spots. Botryodiplodia theobromae was more predominant on blackened seeds. Cephalosporium acremonium, B. theobromae, D. maydis in addition to F. moniliforme were mainly associated with seeds showing discoloured germ ends while wrinkled seeds were observed to harbour F. moniliforme, D. maydis and C. graminicola.

Key words: Diseases, seeds, Zea mays

INTRODUCTION

Maize (Zea mays L.) is the main cereal crop grown in South-western Nigeria. It is used primarily as a staple food for human consumption, animal feeds and as raw material for industrial purposes.

The maize seeds are known to be attacked by various types of pathogens. Of these, fungi account for over 75% of reported cases (Cassini and Cotti 1979). These fungi may damage seeds by causing seed abortion, shrinking the seed, reducing seed size, seed necrosis, seed rot and physiological alteration in seeds (Neergaard 1979; Umechuruba 1986; Shetty 1988). These affect maize seeds either in storage or in the field causing seed discolouration, seed rotting and caking, mycotoxin contamination and loss of viability (Ullstrup 1974; Oyeniran 1977; Pattern 1981). Infected seeds act as media for survival of these fungi as well as their dispersal to disease-free areas (Agarwal and Sinclair 1997). The objective of this study therefore was to isolate, identify and obtain information on the incidence of various seedborne fungi associated with different types of discolouration and abnormalities on seeds of three cultivars of maize common among farmers in Southwestern Nigeria.

MATERIALS AND METHODS

Seeds of three cultivars of maize viz: TZSR-W,

DMRESR-W and DMRLSR-W (Tagged samples 1, 2 and 3, respectively), were obtained from the Institute of Agricultural Research and Training (IAR&T), Ibadan and International Institute of Tropical Agriculture (IITA), Ibadan. maize cultivars were grown in Ibadan, Nigeria. The seeds were allowed to dry on the field, and samples were collected immediately after harvest on 10th and 12th of August 1997 and 1998. These seeds were subjected to visual observation and examination under stereoscopic microscope. Seeds that showed distinct symptoms were selected and categorized into three groups viz: discoloured seeds, wrinkled seeds and seeds with discoloured embryo end. Discoloured seeds were further sub-grouped into seeds with white streaks, seeds with brown spots, seeds with pink/purple discolouration and blackened seeds.

Isolation of fungi from seeds

Infected seeds of each cultivar were surfacesterilized in 2% available chlorine in NaOCI for 15min and then rinsed for 2min each in three changes of sterile distilled water prior to plating. Five seeds were plated in each Petri dish containing 10ml of potato dextrose agar (PDA). In each of the categories and subgroupings, a total of 400 seeds were plated in four replicates of 100 seeds per replicate for each cultivar. These Petri dishes were

²Department of Crop Protection and Environmental Biology, University of Ibadan, Ibadan, Nigeria.

arranged in a Gallenkamp illuminated growth chamber (model 3B5202B) at a temperature of 28 °C under alternating cycles of 12h light and 12h darkness. On the 8th day, incubated seeds were observed for fungal growth and identification under stereoscopic and compound microscopes. Identification was on the basis of the presence and the characteristics of typical fungal structures such as conidia and hyphae (Barnett and Hunter 1972; Benoit and Mathur 1970; Chidambaram et al. 1973).

The number of seeds infected by each kind of fungus was counted and when more than one fungus grew on the same seeds, it was regarded as multiple infection. The data collected were transformed prior to analysis using square root transformation method. Analysis of variance and mean separation were performed using Statistical Analysis Software.

RESULTS

Fusarium moniliforme, Cephalosporium acremonium, and Nigrospora oryzae were associated with white streaks radiating from the embryonic ends of the maize seeds. However, the incidence of F. moniliforme was significantly higher than other fungi in all the three cultivars tested (Table On cultivar TZSR-W, F. moniliforme was identified from 98% of seeds that showed white The incidence of F. moniliforme on seeds that showed purple/pink discolouration was significantly higher than any other fungus isolated on cultivars TZSR-W (89.5%) and DMRLSR-W (52.0%).The percentage incidence of F. graminearum was also significantly high on cultivars DMRESR-W (49.0%) and DMRLSR-W(49.5%). F. semitectum was also isolated although in traces (0.5%) in seeds exhibiting purple/pink discolouration in cultivar DMRLSR-W.

Fusarium moniliforme, Drechslera maydis, Curvularia lunata and Colletotrichum graminicola were observed on seeds with brown spots. incidence of D. maydis (45%) on cultivar TZSR-W

was significantly higher than that of other fungi isolated from brown spotted seeds.

Fusarium moniliforme, D. maydis and Botryodiplodia theobromae were mostly associated with blackened seeds of the three cultivars tested. The percentage incidence of B. theobromae on the blackened seeds was significantly higher than any other observed fungi (Table 1). Table 1 also shows that the incidence of Aspergillus spp was significantly high on the seeds of all the cultivars regardless of the type of discolouration.

Table 2. Percentage incidence of fungi in seeds with discoloured embryonic end and wrinkled seeds of three maize cultivars.

	Seeds with		ured	Wrinkled seeds			
Eurgus	1*	2	3		2	3	
Fusarum moniliforme	88.5a**	69.8a	74.0a	72.8a	56.0a	48.5a	
Cephalosporium acremonium	9.0b	2.5c	0.5c	0.0d	0.0c	0.0c	
Drechslera maydis	11.5b	13.5b	8.0bc	11.3bc	2.5c	9.5b	
Botryodiplodia theobramae	8.0b	9.5bc	6.5bc	10.0bc	0.0c	0.0c	
Colletotrichum graminicola	0.0c	0.5c	2.5bc	7.5c	9.8b	0.5c	
Curvularia lunata	0.0c	0.0c	10.5b	2.0c	0.5c	2.0c	
Aspergillus spp	16.0b	21.5b	18.0b	21.0b	43.0a	46.0a	
Penicillium spp	0.0c	0.0c	0.0c	0.0c	6.8b	12 5b	

¹⁼Cultivar TZSR-W: 2=DMRESR-W, 3=DMRLSR-W

Table 2 shows that the incidence of F. moniliforme was significantly higher than that of C. acremonium, D. maydis, C. graminicola and Aspergillus spp on seeds showing distinct discoloured embryo end in all the three maize cultivars used. Of all the fungi isolated from wrinkled seeds, the percentage incidence of F. moniliforme was the highest. D. maydis, B. theobromae, C. graminicola and Curvularia lunata were other fungi associated with wrinkled seeds. The incidence of Aspergillus spp was however, significantly higher in the varieties 2 and 3.

DISCUSSION

The seed-borne fungi isolated include both field and storage fungi. F. moniliforme, D. maydis, B. theobromae, C. acremonium, and Colletotrichum graminicola were the most important field fungi

Table 1: Seed-borne fungi associated with four different types of discolouration on seeds of three maize cultivars (% incidence)

Eungus	White streaks			Pink/purple			Brown spots			Blackened seeds		
	1*	2	3	1	2	3	ī	2	3	1	2	3
Fusarium moniliforme	98.0a	83.0a	87 Oa	89 5a	38.5b	52.0a	0 0d	18.5b	26.3a	8.5c	26.5b	20.5b
usarium graminearum	0.0c	6.5bc	0.5c	14.0b	49.0a	49.5a	0.0d	0.0d	0.0c	0.0d	0.0c	0.0d
usarium semitectum	0.0c	0.0c	0.0 c	0.0c	0.0d	0.5c	11 Oc	6.5c	2.5c	0.0 d	0.0c	0.0d
Drechslera maydis	0.0c	0.0c	0.0c	0 0c	0.0d	0 Oc	45.8a	10.8bc	6.0bc	15.0c	21.0b	28.3b
Lurvularia lunata	0.0c	0.0c	0.0c	0 Oc	0.0d	0 0c	10 0c	9.5bc	13.8b	0.0d	0.0c	0.0d
Colletotrichum graminicola	0.0c	0.0c	0 Oc	0 0c	0 Od	0 0c	0. 0 d	15.8b	18.5ab	17.8c	0.0c	0. 0 d
'e; halosporium acremonium	5. 5 c	12.5b	10.8b	0.0c	0.0d	0 Oc	0 Od	0.0d	0.0c	0.0d	0.0c	0.0d
Diplodia maydis	0.0c	0.0c	0.0c	0.0c	0 Od	0 Oc	19.0b	0.0d	0.0c	0.0d	0.0c	0.0d
ligrospora oryzae	0.0c	6.0bc	1.5c	0.0c	0.0d	0.0c	0.0d	0.0d	0.0c	0.0d	0.0 c	0.0d
lotryodiplodia theobromae	0.0c	0.0c	0.0c	0.0c	0.0d	0.00	0.0d	0.0d	0.0c	63.0a	49.5a	53.5a
Spergillus spp	21.0b	14.5b	10.36	18.5b	13.3c	16.56	28.5b	35.0a	26.0a	31.0b	17.5b	13.5c
^o enicillium spp	0.5c	1.8c	3.5bc	0 Oc	0.0d	0 0c	0.0d	0.0d	0.0c	0.0d	0.0c	0. 0d

I=cultivar TZSR-W; 2=DMRESR-W, 3=DMRLSR-W

^{**} Each value is a mean of four replicates (100 seeds/replicate/abnormality) Means followed by same letter(s) in the same column are not significantly different at P 0.05 (Duncan's Multiple Range Test).

^{* 1=} COURT (ZON-W, Z=DIMEDIC-W, Z=DIMEDIC-T)

** Each value is a mean of four replicates (100 seeds/replicate/cultivar/abnormality)

Means followed by the same letter in the same coloumn are not significantly different at P 0.05 according to Duncan's Multiple Range Test.

isolated in the study. Other field fungi included Curvularia lunata, F. semitectum and Nigrospora Oryzaae. The storage fungi were Aspergillus spp and Penicillium spp.

The association of *F. moniliforme* with all the different types of seed discolouration and abnormalities, and its high incidence may be due to the susceptibility of the maize crop to attack by this fungus. *Fusarium moniliforme* is known to cause seed and ear rots, stalk rot and leaf spots (Headrick and Pataky 1989). Thomas and Buddenhagen (1980) and Zummo and Scott (1992) have also observed a higher incidence of *F. moniliforme* from maize seeds. In the present study also, this fungus was predominant in the discoloured and wrinkled seeds.

Maize white streaks, often radiating from embryonal end of the seeds, are due to infection by F. moniliforme, C. acremonium and N. oryzae. Although Kumar (1986) observed the association of F. moniliforme and C. acremonoium with white streaks in maize seeds, Nigrospora oryzae is reported eed for the first time. Fusarium moniliforme and F. graminearum was associated with pinkish to purple seeds. A similar observation was reported by Neergaard (1979). Brown spots on maize seeds were associated with F. moniliforme, F. graminearum, D. maydis, C. lunata and F. semitectum. Blackened maize seeds were observed to be infected with B. theobromae, D. maydis, F. moniliforme and C. graminicola. However, Kumar and Shetty (1983), Neergaard (1979;1981) and Singh and Singh (1981) observed that blackening of seeds of maize is usually caused by B. theobromae. Infection levels ranging from 0.5% to 71.5% have been observed on blackened seeds by Singh and Singh (1981). Similar results were obtained in the present study.

Aspergillus and Penicillium spp. were the storage fungi isolated from the different forms of dicolourations and abnormalities. Aspergillus spp were constantly observed in all the different categories of seeds found in all the three cultivars tested. The influence of these fungi on germination and growth of maize plants is, however, negligible (Kumar and Shetty 1983). Some Aspergillus spp. are, however, destructive on stored products and their presence is indicative of poor storage conditions. The findings in this investigation suggest that maize seeds with various forms of discolouration and abnormalities have the potential to cause seed deterioration in storage and epiphytotics in the field.

ACKNOWLEDGMENTS

This work forms part of the M.Phil. thesis submitted to the University of Ibadan by the senior author. The senior author is grateful to the entire staff of the TRIP Pathology Laboratory, International Institute of Tropical Agriculture (IITA), Ibadan for allowing unrestricted use of their facilities.

REFERENCES

- Agarwal UK and Sinclair JB 1997 Principles of seed pathology Vol. 1 and 2. Boca Raton, Fla. CRC Press.
- Barnett HC and Hunter BB 1972 Illustrated genera of imperfect fungi (3rd ed). Burgress Publishing Co. Minneapolis, MN. 20pp.
- Benoit HC and Mathur SB 1970 Identification of species of *Curvularia* on Rice seeds. Proc International Seed Testing Association. 35:99-119.
- Cassini R and Cott I 1979 Parasitic diseases of maize. Ciba Geigy Ltd. pp 72-80. ASP St Minnesota.
- Chidambaram P, Mathur SB and Neergaard P 1973 Identification of seed-borne *Drechslera* species. Friesia. 10:165-207.
- Headrick JM and Pataky J K 1989 Resistance to kernel infection by *Fusarium moniliforme* in unbred lines of sweetcorn and effect of infection on emergence. Plant Disease. 73; 889-892.
- Kumar V 1986 Studies on some seed-borne fungal disease of maize in Karnataka. Ph.D thesis, University of Mysore, India. 199pp.
- Kumar V and Shetty H S 1983 Seed-borne nature and trasmission of *Botryodiplodia thebromae* in maize (*Zea mays*). Seed Science and Technology. 11:781-789.
- Neergaard P 1979 Introduction to methods of seedhealth testing. Seed Science and Technology 7: 601-635.
- Neergaard P 1981 Risk of EPPO Region from seedborne pathogens. EPPO Bulletin. 11:207-212.
- Oyeniran JO 1977 Fungal deterioration of maize during storage in Nigeria. Nigerian Journal of Plant Protection. 3: 102-105.
- Pattern CR 1981 Aflatoxins and disease. American Journal of Tropical Medical Hygiene. 30: 422-425.
- Shetty HC 1988 Different types of damages in seeds caused by seed-borne fungi. Pp 53-62 In: Seed pathology (Eds S.B.Mathur and J.Jorgensen) Proc. of CTA Seminar, Copenhagen, Denmark. 20-25 June 1988.
- Singh DV and Singh T 1981 Location of Fusarium moniliforme in kernels of maize and disease

- transmission. Indian Journal of Physiology and Plant Pathology 7: 32-36.
- Thomas MD and Buddenhagen IW 1980 Incidence and persistence of *Fusarium moniliforme* in symptomless maize kernels and seedlings in Nigeria. Mycologia. 72:882-887.
- Ullstrup AJ 1974 Corn diseases in United States and their control. U.S. Agric Handbook No 199. United States Department of Agriculture,
- Washington D.C. 55pp.
- Umechuruba CI 1986 Effect of thioral on seed-borne fungi associated with maize varieties grown in Eastern Nigeria. Tropical Pest Management 32 (1):27-30.
- Zummo N and Scott GE 1992 Interaction of Fusarium moniliforme and Aspergillus flavus on kernel infection and Aflatoxin contamination in